

FORM PTO-1390 (Modified) (REV 11-2000)		U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE		ATTORNEY'S DOCKET NUMBER <b>03528.0131.PCUS00</b>
TRANSMITTAL LETTER TO THE UNITED STATES DESIGNATED/ELECTED OFFICE (DO/EO/US) CONCERNING A FILING UNDER 35 U.S.C. 371				U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR <b>Not Yet Assigned 09/856341</b>
INTERNATIONAL APPLICATION NO. <b>PCT/DE99/03692</b>	INTERNATIONAL FILING DATE <b>17 November 1999</b>	PRIORITY DATE CLAIMED <b>18 November 1998</b>		
TITLE OF INVENTION <b>DERIVATIZATION OF SUPPORT SURFACES</b>				
APPLICANT(S) FOR DO/EO/US <b>Markus BEIER</b>				
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:				
<p>1. <input checked="" type="checkbox"/> This is a <b>FIRST</b> submission of items concerning a filing under 35 U.S.C. 371.</p> <p>2. <input type="checkbox"/> This is a <b>SECOND</b> or <b>SUBSEQUENT</b> submission of items concerning a filing under 35 U.S.C. 371.</p> <p>3. <input type="checkbox"/> This is an express request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (24) indicated below.</p> <p>4. <input checked="" type="checkbox"/> The US has been elected by the expiration of 19 months from the priority date (Article 31).</p> <p>5. <input checked="" type="checkbox"/> A copy of the International Application as filed (35 U.S.C. 371 (c) (2))        a. <input checked="" type="checkbox"/> is attached hereto (required only if not communicated by the International Bureau).        b. <input type="checkbox"/> has been communicated by the International Bureau.        c. <input type="checkbox"/> is not required, as the application was filed in the United States Receiving Office (RO/US).</p> <p>6. <input checked="" type="checkbox"/> An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).        a. <input checked="" type="checkbox"/> is attached hereto.        b. <input type="checkbox"/> has been previously submitted under 35 U.S.C. 154(d)(4).</p> <p>7. <input type="checkbox"/> Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c)(3))        a. <input type="checkbox"/> are attached hereto (required only if not communicated by the International Bureau).        b. <input type="checkbox"/> have been communicated by the International Bureau.        c. <input type="checkbox"/> have not been made; however, the time limit for making such amendments has NOT expired.        d. <input type="checkbox"/> have not been made and will not be made.</p> <p>8. <input type="checkbox"/> An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).</p> <p>9. <input type="checkbox"/> An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).</p> <p>10. <input type="checkbox"/> An English language translation of the annexes of the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).</p> <p>11. <input checked="" type="checkbox"/> A copy of the International Preliminary Examination Report (PCT/IPEA/409).</p> <p>12. <input checked="" type="checkbox"/> A copy of the International Search Report (PCT/ISA/210).</p>				
<b>Items 13 to 20 below concern document(s) or information included:</b>				
<p>13. <input type="checkbox"/> An Information Disclosure Statement under 37 CFR 1.97 and 1.98.</p> <p>14. <input type="checkbox"/> An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.</p> <p>15. <input checked="" type="checkbox"/> A <b>FIRST</b> preliminary amendment.</p> <p>16. <input type="checkbox"/> A <b>SECOND</b> or <b>SUBSEQUENT</b> preliminary amendment.</p> <p>17. <input type="checkbox"/> A substitute specification.</p> <p>18. <input type="checkbox"/> A change of power of attorney and/or address letter.</p> <p>19. <input type="checkbox"/> A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 35 U.S.C. 1.821 - 1.825.</p> <p>20. <input type="checkbox"/> A second copy of the published international application under 35 U.S.C. 154(d)(4).</p> <p>21. <input type="checkbox"/> A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).</p> <p>22. <input checked="" type="checkbox"/> Certificate of Mailing by Express Mail</p> <p>23. <input checked="" type="checkbox"/> Other items or information:</p>				
<b>Clean Version of All Pending Claims; Clean Version of Amended Paragraph; return receipt postcard</b>				

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR, No Y Assigned) <b>09/856341</b>	INTERNATIONAL APPLICATION NO. <b>PCT/DE99/03692</b>	ATTORNEY'S DOCKET NUMBER <b>03528.0131.PCUS00</b>
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24. The following fees are submitted:

**BASIC NATIONAL FEE ( 37 CFR 1.492 (a) (1) - (5) ) :**

- |  |           |
|--|-----------|
| <input type="checkbox"/> Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO ..... | \$1000.00 |
| <input checked="" type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO .....  | \$860.00  |
| <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO .....  | \$710.00  |
| <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) paid to USPTO but all claims did not satisfy provisions of PCT Article 33(1)-(4) .....   | \$690.00  |
| <input type="checkbox"/> International preliminary examination fee (37 CFR 1.482) paid to USPTO and all claims satisfied provisions of PCT Article 33(1)-(4) .....   | \$100.00  |

**CALCULATIONS PTO USE ONLY**

**ENTER APPROPRIATE BASIC FEE AMOUNT =**

**\$860.00**

Surcharge of **\$130.00** for furnishing the oath or declaration later than  20  30 months from the earliest claimed priority date (37 CFR 1.492 (e)).

**\$130.00**

CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	
Total claims	20 - 20 =	0	x \$18.00	<b>\$0.00</b>
Independent claims	3 - 3 =	0	x \$80.00	<b>\$0.00</b>

Multiple Dependent Claims (check if applicable).

**TOTAL OF ABOVE CALCULATIONS = \$1,260.00**

Applicant claims small entity status. (See 37 CFR 1.27). The fees indicated above are reduced by 1/2.

**\$630.00**

**SUBTOTAL = \$630.00**

Processing fee of **\$130.00** for furnishing the English translation later than  20  30 months from the earliest claimed priority date (37 CFR 1.492 (f)).

**\$0.00**

**TOTAL NATIONAL FEE = \$630.00**

Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable).

**\$0.00**

**TOTAL FEES ENCLOSED = \$630.00**

	<b>Amount to be: refunded</b>	<b>\$</b>
	<b>charged</b>	<b>\$</b>

- A check in the amount of **\$630.00** to cover the above fees is enclosed.
- Please charge my Deposit Account No. \_\_\_\_\_ in the amount of \_\_\_\_\_ to cover the above fees. A duplicate copy of this sheet is enclosed.
- The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. **08-3038** A duplicate copy of this sheet is enclosed.
- Fees are to be charged to a credit card. **WARNING: Information on this form may become public. Credit card information should not be included on this form.** Provide credit card information and authorization on PTO-2038.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO:

Albert P. HALLUIN  
HOWREY SIMON ARNOLD & WHITE, LLP  
301 Ravenswood Avenue  
Box 34  
Menlo Park, CA 94025

**SIGNATURE**

**Albert P. Halluin, Viola T. Kung**

**NAME**

**25,227, 41,131**

**REGISTRATION NUMBER**

**May 18, 2001**

**DATE**

## IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:

Beier, M.

Group Art Unit: To Be Assigned

Application Serial No.: To Be Assigned

Filed: May 18, 2001

Examiner: To Be Assigned

For: **DERIVATIZATION OF SUPPORT  
SURFACES**

Attorney's Docket No.:

03528.0131.PCUS00

PRELIMINARY AMENDMENTCommissioner for Patents  
Washington, D.C. 20231

Sir:

Applicants are submitting herewith a Preliminary Amendment. The Examiner is respectfully requested to enter the amendments prior to considering the application.

THE AMENDMENTIn the Specification:

Page 1, line 2, before "The present invention" insert --This application is a National Stage of International Application PCT/DE99/03692, filed November 17, 1999; which claims the priority of DE 198 53 242.3 filed November 18, 1998--.

In the Claims

3. (Amended) The method according to claim [1 or] 2, wherein the functional group is an amine, hydroxyl, phosphate, carboxyl, carbonyl, thiol or amide group.
  
4. (Amended) The method according to [any one of claims 1 to 3] claim 2, wherein the activating reagent is acryloylchloride, 4-nitrophenylchloroformate, carbonyl diimidazole, phenyl chloroformate, phosgene, disphosgene, triphosgene, EDC(N-(3-dimethylaminopropyl)-N'-ethyl-carbodiimide hydro-

- chloride), N,N'-diisopropyl carbodiimide, dicyclohexyl carbodiimide, disuccinimidyl carbonate, disuccinimidyl oxalate, dimethylsuberimidate dihydrochloride or phenylene diisothiocyanate.
5. (Amended) The method according to [any one of claims 1 to 4] claim 1, wherein the amine component is selected from the group consisting of monoamines, bis-amines or polyamines.
9. (Amended) The method according to [any one of claims 1 to 8] claim 1, wherein the steps of the reaction with an activating reagent and an amine component are carried out several times.
11. (Amended) The method according to [any one of the preceding claims] claim 1, wherein a positive charge is built up in controlled fashion on the support surface.
12. (Amended) The method according to [any one of claims 1 to 11] claim 2, wherein the support surface derivatized according to [any one of claims 1 to 11] claim 2 is additionally activated prior to the attachment of biopolymers.
13. (Amended) The method according to claim 12, wherein said activating agent is disuccinimidyl carbonate, disuccinimidyl oxalate, glutaraldehyde, dimethylsuberimidate dihydrochloride or phenylene diisothiocyanate [are used as activation agent].
15. (Amended) [The] A support suitable for the attachment of biopolymers, which includes linkers in the form of dendrimer structures on its surface.

Cancel claim 16.

17. (Amended) [Use according to claim 16] The method according to claim 18 or 19, wherein the biopolymers are selected from the group consisting of DNA, RNA, nucleotide analogs, peptides, proteins or antibodies.

Add new claims 18-19.

--18. (New) A method for attaching biopolymers to a support comprising the step of binding biopolymers to the support surface derivatized and activated according to claim 12.

19. (New) A method for attaching biopolymers to a support comprising the step of binding biopolymers to the support surface derivatized and activated according to claim 14 or 15.--

**REMARKS**

**The Amendment**

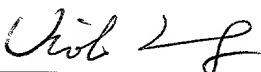
The above amendments correct the improper format of multiple dependent claims.

New claims 18 and 19 are supported by claim 16 as filed.

No new matter is added in any of the amendments. The Examiner is respectfully requested to enter all the amendments.

Respectfully submitted,

Date: May 18, 2001

  
\_\_\_\_\_  
Albert P. Halluin, Reg. No. 25,227  
Viola T. Kung, Reg. No. 41,131

**HOWREY & SIMON**  
301 Ravenswood Avenue  
Box No. 34  
Menlo Park, CA 94025  
(650) 463-8109

**CLEAN VERSION OF AMENDED PARAGRAPH**

**In the Specification:**

This application is a National Stage of International Application PCT/DE99/03692, filed November 17, 1999; which claims the priority of DE 198 53 242.3 filed November 18, 1998.

**CLEAN VERSION OF ALL PENDING CLAIMS**

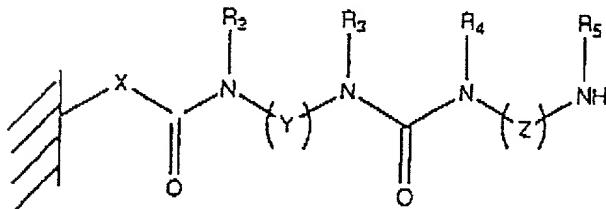
1. A method of derivatizing carriers or supports, wherein a functional group is activated on a carrier or support surface by reaction with an activating reagent and subsequently reacted with an amine component.
2. The method according to claim 1, wherein the supports are selected from the group consisting of glass, sheets or films or membranes made from polypropylene, nylon, cellulose, cellulose derivatives (e.g. cellulose acetate, cellulose-mixed ester), polyether sulfones, polyamides, polyvinyl chloride, polyvinylidene fluoride, polyester, polyethylene or Teflon.
3. The method according to claim 2, wherein the functional group is an amine, hydroxyl, phosphate, carboxyl, carbonyl, thiol or amide group.
4. The method according to claim 2, wherein the activating reagent is acryloylchloride, 4-nitrophenylchloroformate, carbonyl diimidazole, phenyl chloroformate, phosgene, disphosgene, triphosgene, EDC(N-(3-dimethylaminopropyl)-N'-ethyl-carbodiimide hydrochloride), N,N'-diisopropyl carbodiimide, dicyclohexyl carbodiimide, disuccinimidyl carbonate, disuccinimidyl oxalate, dimethylsuberimidate dihydrochloride or phenylene diisothiocyanate.
5. The method according to claim 1, wherein the amine component is selected from the group consisting of monoamines, bis-amines or polyamines.
6. The method according to claim 5, wherein the monoamine is 2-aminoethanol, 6-amino-1-hexanol, 2-(4-aminophenyl)ethanol, 5-amino-n-valeric acid, 2-(2-aminoethoxy)ethanol or 3-amino-1,2-propanediol.
7. The method according to claim 5, wherein the bis-amine is 1,4-bis(3-aminopropoxy)butane, O,O'-bis(2-aminopropyl)polyethylene glycol 500 (=

Jeffamine 500), O,O'-bis(2-aminopropyl)polyethylene glycol 130 (= Jeffamine 130), 4,7,10-trioxa-1,13-tridecaneamine or ethylene diamine.

8. The method according to claim 5, wherein the polyamine is tetraethylene pentamine, spermine, spermidine, 4,7,10-trioxa-1,13-tridecanediamine or 4-aminomethyl-1,8-octanediamine.
9. The method according to claim 1, wherein the steps of the reaction with an activating reagent and an amine component are carried out several times.
10. The method according to claim 9, wherein dendrimer structures are built up on the support surface.
11. The method according to claim 1, wherein a positive charge is built up in controlled fashion on the support surface.
12. The method according to claim 2, wherein the support surface derivatized according to claim 2 is additionally activated prior to the attachment of biopolymers.
13. The method according to claim 12, wherein said activating agent is disuccinimidyl carbonate, disuccinimidyl oxalate, glutaraldehyde, dimethylsuberimidate dihydrochloride or phenylene diisothiocyanate.
14. A support suitable for the attachment of biopolymers, wherein the surface of the support includes linkers having the following structure:

X = O, NHR<sub>1</sub>  
Y, Z = may be equal or different and be selected from  
-(CH<sub>2</sub>)<sub>n</sub>-  
-(CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>-CH<sub>2</sub>CH<sub>2</sub>  
-(CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>-CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>-, etc.  
R<sub>1</sub>-R<sub>5</sub> = may be equal or different and be selected from a straight-chain or branched alkyl residue having 1 to 30 carbon atoms, straight-chain or branched alkenyl residue having 2 to 30 carbon atoms, monocyclic or polycyclic alkyl residue having 3 to 30 carbon atoms or monocyclic or polycyclic alkenyl residue having 4 to 30 carbon atoms or monocyclic or polycyclic residue having 6 to 30 carbon atoms, the residues being optionally substituted by one or several substituents (e.g. OH, carboxyl, carbonyl, phosphate)  
n = 1 to 50

or



X = O, NHR<sub>1</sub>  
Y, Z = may be equal or different and be selected from  
-(CH<sub>2</sub>)<sub>n</sub>-  
-(CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>-CH<sub>2</sub>CH<sub>2</sub>  
-(CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>-CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>-, etc.  
R<sub>1</sub>-R<sub>5</sub> = may be equal or different and be selected from a straight-chain or branched alkyl residue having 1 to 30 carbon atoms, straight-chain or branched alkenyl residue having 2 to 30 carbon atoms, monocyclic or polycyclic alkyl residue having 3 to 30 carbon atoms or monocyclic or polycyclic alkenyl residue having 4 to

30 carbon atoms or monocyclic or polycyclic residue having 6 to 30 carbon atoms, the residues being optionally substituted by one or several substituents (e.g. OH, carboxyl, carbonyl, phosphate)

n = 1 to 50

15. A support suitable for the attachment of biopolymers, which includes linkers in the form of dendrimer structures on its surface.
17. The method according to claim 18 or 19, wherein the biopolymers are selected from the group consisting of DNA, RNA, nucleotide analogs, peptides, proteins or antibodies.
18. A method for attaching biopolymers to a support comprising the step of binding biopolymers to the support surface derivatized and activated according to claim 12.
19. A method for attaching biopolymers to a support comprising the step of binding biopolymers to the support surface derivatized and activated according to claim 14 or 15.

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Derivatization of Support Surfaces

The present invention relates to a method of derivatizing support or carrier surfaces and to support or carrier surfaces derivatized thereby.

The binding of biopolymers, in particular nucleic acids, to solid support or carrier surfaces is generally obtained to date by the following alternatives:

- 1) Applying biopolymers (e.g. nucleic acids) to surfaces:  
Here above all poly-L-lysine coated glass supports and nylon membranes are used. In this case, the biopolymers are bound to the support by means of charging. When poly-L-lysine coated glass supports are used, it is disadvantageous that no covalent linkage takes place between the coated surface and the biopolymer. The support can only be used once. Furthermore, there are virtually no possibilities of optimizing the distance between biopolymer and support. When nylon membranes are used, it is disadvantageous for the biopolymers to be also largely bound only by means of charging. Although the support may be used several times, it is not possible to optimize the distance between biopolymer and support.
  
- 2) *In situ* structure of biopolymers (e.g. nucleic acids) on surfaces: Here common linker systems are used which originate from the biopolymer synthesis on porous CPG materials. The linker molecules used are usually polyethylene glycol, in particular tetra- or

hexaethylene glycol. The linker molecules are usually applied by cost-intensive reagents in analogy to the phosphoramidite chemistry. Mass production and the application of charges are unfortunately not possible.

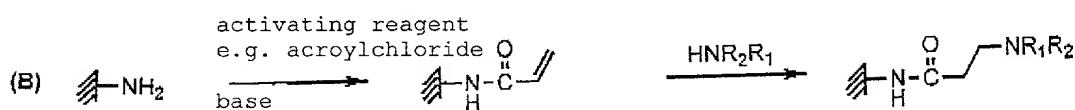
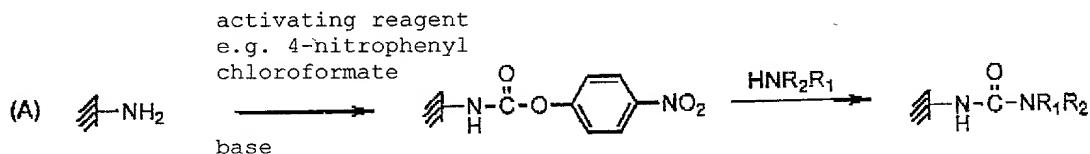
All of these methods also have the drawback of lacking flexibility, and the biopolymers can only be applied on the surface in very limited numbers.

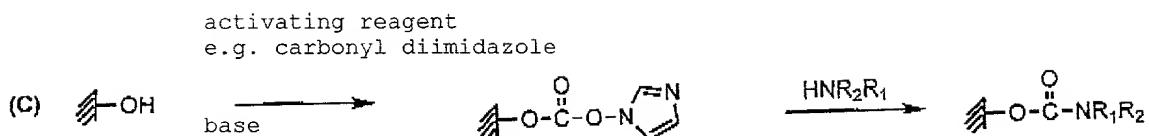
The object of the present invention now consists in providing a method permitting optimum binding of a large number of biopolymers to support surfaces. The object also consists in providing a support surface whose binding capacity has been increased many times by carrying out a surface modification.

These objects are achieved by the subject matters defined in the claims.

In particular, this object is achieved by a method with which a functional group is activated on a support surface by an activating reagent and is subsequently reacted with an amine component.

The method according to the invention is preferably based on the following synthesis principles:





wherein R<sub>1</sub> and R<sub>2</sub> may be equal or different. The meanings of R<sub>1</sub> and R<sub>2</sub> are not subject to limitation and may be H or any organic residue (e.g. a straight-chain or branched alkyl residue having 1 to 30 carbon atoms, a straight-chain or branched alkenyl residue having 2 to 30 carbon atoms, a monocyclic or polycyclic alkyl residue having 3 to 30 carbon atoms or a monocyclic or polycyclic alkenyl residue having 4 to 30 carbon atoms or a monocyclic or polycyclic residue having 6 to 30 carbon atoms, wherein the residues may optionally be substituted by one or more substituents (e.g. OH, carboxyl, carbonyl, phosphate)). The base may be any basic compound, e.g. diisopropylethylamine.

Any straight-chain or branched C<sub>1-30</sub> alkyl residue may be used. Examples thereof are methyl, ethyl, propyl, isopropyl, butyl, isobutyl, tert.-butyl, n-butyl, n-hexyl, 2-methylpentyl, 2,3-dimethylbutyl, n-heptyl, 2-methylhexyl, 2,3-dimethylpentyl, 3,3-dimethylpentyl, 3-ethylpentyl, n-octyl, 2,2-dimethylhexyl, 3,3-dimethylhexyl, 3-methyl-3ethylpentyl groups. Short alkyl chains, such as methyl, ethyl, propyl and isopropyl groups, are preferred.

Any straight-chain or branched C<sub>2-30</sub> alkenyl residue may be used. Examples thereof are vinyl, propenyl, isopropenyl, allyl, 2-methylallyl, butenyl or isobutenyl, hexenyl or isohexenyl, heptenyl or isoheptenyl, octenyl or isoctenyl

groups. Vinyl, propenyl and isopropenyl groups are preferred.

The cycloalkyl residue having 3 to 30 carbon atoms may be any cycloalkyl residue. Examples thereof are cyclopropyl, cyclobutyl, cyclopentyl or cyclohexyl, cycloheptyl, cyclooctyl, cyclononyl or cyclodecyl group. Cyclopropyl, cyclobutyl, cyclopentyl and cyclohexyl groups are preferred.

The cycloalkenyl residue having 4 to 30 carbon atoms may be any cycloalkenyl residue. Examples thereof are cyclobutenyl, cyclopentenyl or cyclohexenyl, cycloheptenyl, cyclooctenyl, cyclononenyl or cyclodecencyl groups. Cyclobutenyl, cyclopentenyl, or cyclohexenyl groups are preferred.

Examples of polycyclic alkyl or alkenyl residues comprise norbornane, adamantane or benzvalene.

R<sub>1</sub> and R<sub>2</sub> may also be any monocyclic or polycyclic C6-30 aryl residues. Examples thereof are a carbocyclic, monocyclic residue, e.g. the phenyl group, a heterocyclic, monocyclic residue, e.g. the groups thienyl, furyl, pyranyl, pyrrolyl, imidazolyl, pyrazolyl, pyridyl, pyrazinyl, pyrimidinyl, pyrazinyl, thiazolyl, oxazolyl, furazannyl, pyrrolinyl, imidazolinyl, pyrazolinyl, thiazolinyl, triazolyl, tetrazolyl, and the positional isomers of the hetero atom or atoms which may comprise these groups.

By the method according to the invention more or less cross-linked linker systems form on the support surface, which permit binding of an optionally large number of biopolymers on the support surface. Preferred biopolymers are DNA, RNA, nucleic acid analogs, peptides, proteins, antibodies, etc.

According to the invention a functional group is understood to mean a chemical grouping located on a support surface,

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such as an amino group, hydroxyl group, carboxyl group, carbonyl group, thiol, amide or phosphate group.

According to the invention any support or matrix common in this field can be used. They comprise in particular glass, sheets or films or membranes made of polypropylene, nylon, cellulose, cellulose derivatives (e.g. cellulose acetate, cellulose mixed ester), polyethersulfones, polyamides, polyvinyl chloride, polyvinylidene fluoride, polyester, teflon or polyethylene.

0 92 255 17 0 92 255 17

According to the invention an activating reagent is understood to mean a reagent which places functional groups located on the support surface in a condition ready for linkage. Preferred activating reagents are 4-nitrophenyl chloroformate (= chloroformic acid-4-nitrophenylester), carbonyl diimidazole, acryloylchloride (acrylic acid chloride), phenylchloroformate, phosgene, diphosgene, triphosgene, EDC (N-(3-dimethylaminopropyl)-N'-ethylcarbodiimide hydrochloride), N,N'-diisopropyl carbodiimide, dicyclohexyl carbodiimide, disuccinimidyl carbonate, disuccinimidyl oxalate, dimethylsuberimidate dihydrochloride or phenylene diisothiocyanate, the three former ones being most preferred.

According to the invention an amine component is understood to mean monoamines, bis-amines or polyamines. Preferred monoamines are 2-aminoethanol, 6-amino-1-hexanol, 2-(4-aminophenyl)ethanol, 5-amino-n-valeric acid, 2-(2-aminoethoxy)ethanol, 3-amino-1,2-propanediol; preferred bis-amines are 1,4-bis(3-aminopropoxy)butane, O,O'-bis(2-aminopropyl)polyethylene glycol 500 (= Jeffamine 500), O,O'-bis(2-aminopropyl)polyethylene glycol 130 (= Jeffamine 130), 4,7,10-trioxa-1,13-triadecaneamine, ethylene diamine, N-

methylimidazole, diisopropylethylamine; and preferred polyamine are tetraethylene pentamine, spermine, spermidine, 4,7,10-trioxa-1,13-tridecane diamine, 4-aminomethyl-1,8-octane diamine. By incorporating amines preferably positive charges can be implemented into the linker system, since they are present in protonated condition within the physiological region. The degree of the surface charge can be controlled by selecting the corresponding amine. Positive charges effect an easier approach of negatively charged biopolymers (e.g. nucleic acids) and thus facilitate hybridizations on the support surfaces treated according to the invention. The use of bis-amines or polyamines serves for controlling the chain length, i.e. the length of the linker system, by the length of the amine and the number of repeated passing through the below synthesis principles. By selecting the amine it is possible to control the individual character of the linker system, i.e. whether it is rather hydrophobic or hydrophilic. By selecting the amine, other functional groups (e.g. hydroxyl, phosphate, carboxyl, carbonyl groups) may, of course, also be presented on the surface in addition to amine groupings. For example, it is advantageous for introducing a linker carrying OH groups to carry out the reaction with an amine alcohol. When a bifunctional amine is used, the chain is extended linearly. Branchings are incorporated into the linker system by polyfunctional amine reagents. As a result, what is called dendrimer structures are built up (see figures 1 and 2). According to the invention a dendrimer structure means that structures result which start from a defined starting point and have more than one branches. The advantage of this is above all that in the case of support materials with otherwise low charging capacity (e.g. glass) the charging can be increased in well-calculated manner and thus greater amounts of biopolymers can be applied. When polyfunctional

amines ( $m =$  number of the amino functions) are used,  $x$  positions (functions) may be utilized for a binding of biopolymers after  $n$  rounds (1 round = cycle from activation and reaction with amine):

$x = (m-1)^n$  (in the case of activation with 4-nitrophenyl chloroformate or carbonyl diimidazole)

$x = m^n$  (in the case of activation with acryloylchloride)

Arithmetical example: tetraethylene pentamine ( $m = 5$ ), 3 rounds ( $n = 3$ ), acryloylchloride as activating agent ---->  $5^3 = 125$ , i.e. 125 times the charging capacity of the support

In order to obtain a more or less cross-linked linker system on the support surface, the above reactions are repeated in well-calculated fashion. By this, the distance between support material and the biopolymer to be bound is controlled and also the physical properties of the linker are defined. By the method according to the invention a flexible linker system is created. It is flexible in so far as for any use (e.g. binding of DNA, RNA, proteins, antibodies) perfectly prepared support surfaces are provided. In particlar, the method according to the invention is adapted to provide surfaces (supports) suitable for the DNA/biochip technology. The degree of positive charge on the surface can be controlled by a number of protonatable amine functions, above all by the incorporation of polyamines. For example, the objective of increasing the charging density is preferably realized by incorporating branching sites (e.g. polyamines); the increase in the positive surface charge may be effected by the incorporation

of protonatable amino functions, for example; variation of the distance between biomolecule and surface can be controlled by using amines having different chain lengths.

The method according to the invention for modifying the surface is characterized by the following steps. The support materials to be derivatized are admixed in a reaction vessel with 5-100 ml (depending on the size), preferably 10-30 ml, most preferably 20 ml, anhydrous solvent (e.g. dichloroethane, tetrachlorocarbon, THF, DMF, DMSO, HMPT (= HMPA), dichloroethane, acetonitrile) with about 0.5 to 5 mmol, preferably 1-3 mmol, most preferably 1 mmol, activating reagent (e.g. acryloylchloride, 4-nitrophenylchloroformate, carbonyldiimidazole). The reaction is started in the case of acryloylchloride and 4-nitrophenyl chloroformate by adding about 0.5 to 5 mmol, preferably 1-3 mmol, most preferably 1 mmol, diisopropylethylamine (or another non-nucleophilic base, such as triethylamine, pyridine, collidine, lutidine or triisopropylamine). After 1 to 4 hours, preferably 2 hours, of swirling on a shaker, the supports are thoroughly washed with an anhydrous solvent (e.g. dichloroethane, tetrachlorocarbon, etc.) and allowed to dry, preferably air-blown using nitrogen. In order to check the reaction, a quality control is carried out. For example, the above-mentioned aminated PP control piece is subjected to the below described bromophenol blue test. If as compared to a non-reacted sample no blue staining can be detected, the activation was successful. If blue staining occurs, the reaction is repeated. The then activated support materials are admixed in about 5-100 ml (depending on the size), preferably 10-50 ml, most preferably 20 ml, of an amine-free and anhydrous solvent (e.g. DMF, acetonitrile, THF, DMSO, HMPT (HMPA) or dichloromethane) with about 0.5-5 mmol, preferably 1-3 mmol, preferably 1 mmol, amine component and

swirled for about 5-20 hours, preferably overnight. Washing is carried out thoroughly, e.g. in succession with DMF, methanol and dichloroethane, and the then derivatized supports are allowed to dry or are air-blown using nitrogen. In the case of supports activated using acryloylchloride, a reaction time extended to 1.5 to 2.5 days, preferably 2 days, is preferred. For checking the reaction, a quality control is carried out, e.g. the above-mentioned PP control piece is subjected to the below described bromophenol blue test. If as compared to an activated sample of the last step blue staining can be detected, the reaction was successful. If no or only very weak blue staining occurs, the reaction is repeated. The once derivatized supports can already be used as such or be subjected to the described cycle once or several times.

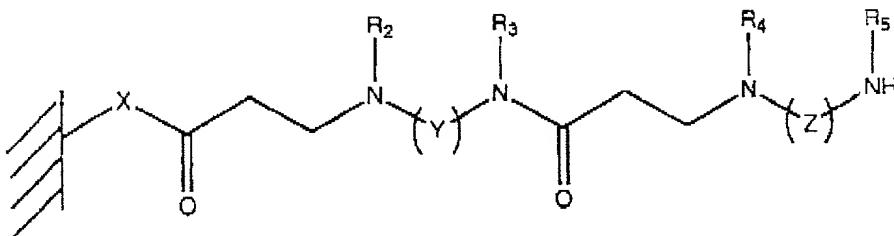
For example, a method based on the blue staining of amino functions by bromophenol blue is suitable for controlling the quality of the support derivatization, i.e. detection takes place via the disappearance or appearance of the blue staining occurring with the activation or reaction with amines. For this purpose, each reaction vessel is provided in addition to the support materials to be derivatized with a piece of aminated polypropylene sheet (PP control piece) which serves for checking, on behalf of all of the support materials in the reaction vessel, the success of the respective reaction step. For this purpose, a piece of the control strip is removed after each reaction and swirled in a 1 % bromophenol blue solution in amine-free DMF and then washed with ethanol. Amino functions and other basic groups yield an intense adsorptive blue staining. After carrying out the activation reactions (e.g. with acryloylchloride, 4-nitrophenylchloroformate, carbonyldiimidazole), detection is made for the disappearance of the blue staining. A

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quantification of the surface charge is possible by means of U.V. spectroscopy. Here, the adsorptive blue staining is removed from the support e.g. by means of a 10 % piperidine solution in DMF and is measured by means U.V. spectroscopy.

The linkers on the support surface have linear structures or dendrimer structures. The structures are built up as follows:

- preferred structure after activation using acryloylchloride



X = O, NHR<sub>1</sub>

Y, Z = may be equal or different and may be selected from

-(CH<sub>2</sub>)<sub>n</sub>-

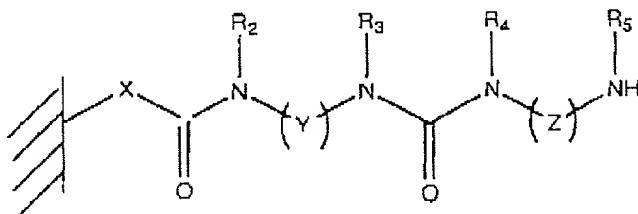
-(CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>-CH<sub>2</sub>CH<sub>2</sub>

-(CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>-CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>-, etc.

R<sub>1</sub>-R<sub>5</sub> = may be equal or different analogous to R<sub>1</sub> and R<sub>2</sub> above

N = 1-50, preferably 1-20, most preferably 1-10

- preferred structure after activation with chloroformic acid nitrophenylester or carbonyldiimidazole



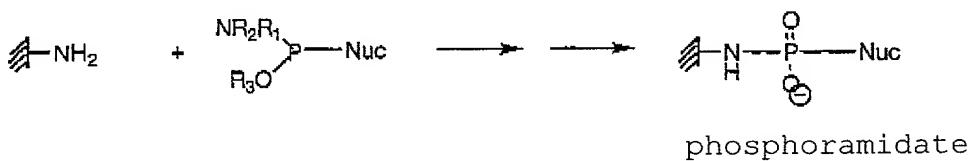
$X = O, NHR_1$   
 $Y, Z =$  may be equal or different and may be selected from  
 $-(CH_2)_n-$   
 $-(CH_2CH_2O)_n-CH_2CH_2$   
 $-(CH_2CH_2CH_2O)_n-CH_2CH_2CH_2-$ , etc.  
 $R_1-R_5 =$  may be equal or different analogous to  $R_1$  and  $R_2$  above  
 $n = 1-50$ , preferably 1-20, most preferably 1-10

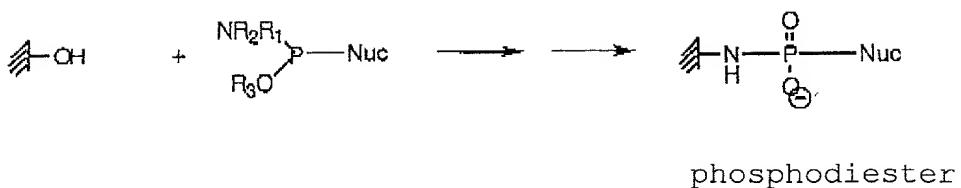
When biopolymers are fixed to support surfaces derivatized with the linker system, a distinction can, in principle, be made between the 2 cases (a) and (b):

- (a) *In situ* synthesis of biopolymers (using the oligonucleotide synthesis as an example):

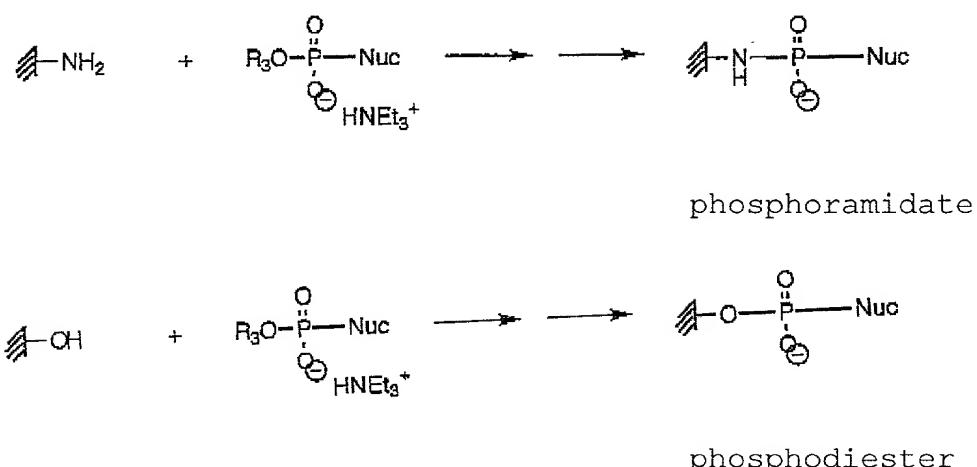
The derivatized support surfaces having terminal amino or hydroxyl functions do not require any further modification for binding the 1<sup>st</sup> oligonucleotide building block according to the phosphoramidite method or also the phosphotriester method. Having reacted the 1<sup>st</sup> nucleotide reagent with the aminated surface, phosphoramidate type bonds are produced. Hydroxylated surfaces result in phosphodiester type bonds.

Phosphoramidite method:





Phosphotriester method:



(Nuc = nucleoside)

R<sub>1</sub> - R<sub>3</sub> = analogous to R<sub>1</sub> and R<sub>2</sub> above

If phosphodiester type bonds are aimed at, it suffices not to react bis-amine in the last step but to react an amino alcohol within the linker-system synthesis.

(b) Binding of "prefabricated" biopolymers:

The binding of the biopolymer provides either the generation of a covalent (permanent) chemical linkage of the biomolecule to the support surface or also a permanent linkage via electrostatic interaction. In the former case, a chemical bond must be produced between the biomolecule and

the surface. In order to produce this bond, e.g. the following procedure is in consideration:

- (A) addition of a promoter
- (B) activation of one of the two reactants (biomolecule or surface). However, an activation of the support surface has to be preferred over an activation of the biomolecule, since undesired cross-reaction among the activated biomolecules cannot be excluded - due to the usually polyfunctional character of the biomolecule - when the biomolecule per se is activated. This activation of the surface is preferably carried out using cross-linkers.

When the biopolymer is bound by electrostatic interaction, the positive charge, which forms by the described linker system using amine-produced dendimeric surfaces, in particular by the incorporation of the amines (preferably bis-amines and polyamine), permits a permanent immobilization of negatively charged biopolymers (e.g. nucleic acids). No further activation of the surface is necessary for this purpose. According to the invention the biopolymer is applied directly onto the dendimeric surface and is fixed permanently. The properties for the permanent immobilization of biopolymers by electrostatic interaction are increased according to the invention by alkylating, e.g. methylating, the dendimeric surface beforehand.

#### (A) Binding of the biopolymer using a promoter

The possibility of using a promoter (catalyst) for producing the covalent bond between surface and biomolecule, is substantially a 3-component reaction. If as in the present invention possibilities are aimed at which shall permit to fix in highly parallel manner a large number of biopolymers to accurately predetermined positions on a support, this can be done due to cross-contamination only with more difficulty

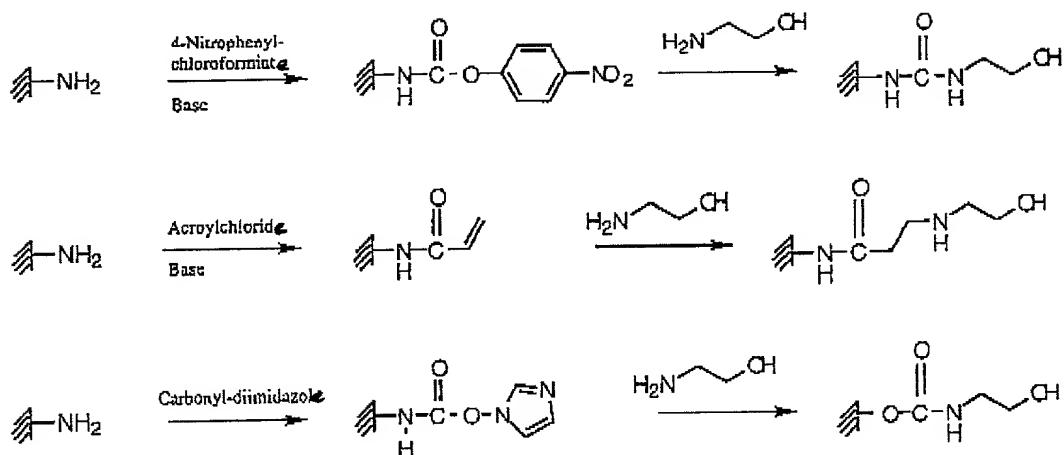
on plane surfaces using a 3-component system (sheets, glass or the like). Therefore, the promoter method is a special case for the covalent linkage of biopolymers on membrane type surfaces. Only here can the promoter be provided locally to the biomolecule to be bound in the pores of the membrane without cross-contamination having to be expected (cf. Example 4). For example, reagents of the carbodiimide type, such as diisopropylcarbodiimide, EDC or DCC (dicyclohexylcarbodiimide), are suitable as promoters. Furthermore, the terminal groups should on the support and the biomolecule be preferably orthogonal relative to one another, i.e. they should be able to enter into a useful chemical bond. Here, preferably the following combinations of the terminal groups to be reacted result:

<u>Support surface</u>	<u>biomolecule</u>	<u>resulting binding type</u>
Amino (-NHR)	carboxy (-COOH)	amide bond
Amino (-NHR)	phosphate (-PO <sub>4</sub> <sup>2-</sup> )	phosphoramidite
Hydroxyl (-OH)	carboxy (-COOH)	ester
Hydroxyl (-OH)	phosphate (-PO <sub>4</sub> <sup>2-</sup> )	phosphodiester
Carboxy (-COOH)	amino (-NHR)	amide bond
Carboxy (-COOH)	hydroxyl (-OH)	ester
Phosphate (-PO <sub>4</sub> <sup>2-</sup> )	amino (-NHR)	phosphoramidate
Phosphate (-PO <sub>4</sub> <sup>2-</sup> )	hydroxyl (-OH)	phosphodiester

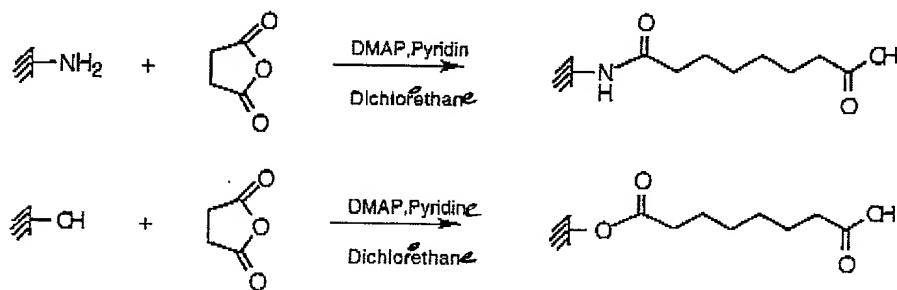
Since the reactive groups (amine, hydroxyl, phosphate or carboxyl) are more or less given by the biomolecule, it is recommended that every biomolecule type has available a corresponding orthogonal surface type. In order to make available the support surface derivatization with the linker system for all of the above-mentioned combinations of terminal groups, methods were developed for re-derivatizing the surface of terminal amino functions (or also hydroxyl)

on terminal hydroxyl, carboxyl and phosphate groups (cf. Examples 5, 6, 7).

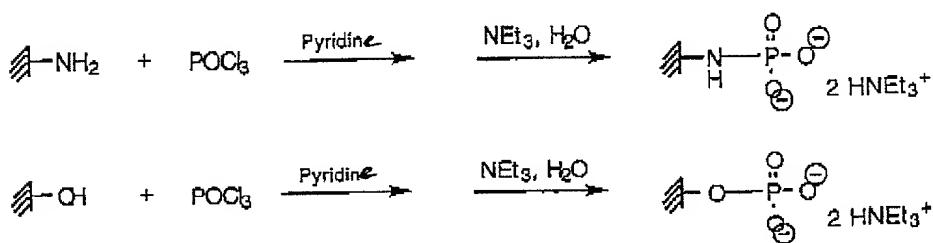
Re-derivatization on terminal hydroxyl:



Re-derivatization on terminal carboxyl:



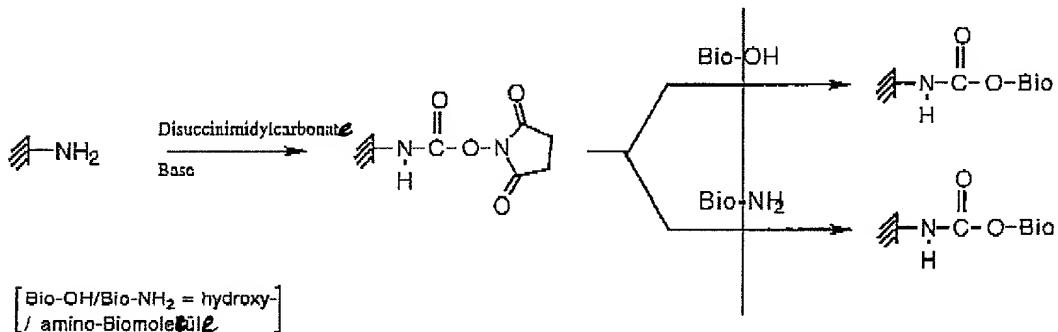
Re-derivatization on terminal phosphate:



(B) Binding of the biopolymer by means of cross-linkers

A covalent surface activation is understood to mean an activation in which cross-linkers are so to speak grafted as a terminal group onto the linker system built up according to the invention beforehand (flexible linker system). They are above all activated ester, aldehyde, imidoester or isothiocyanate functions which are capable of covalently binding without further chemical step to biopolymers dissolved in aqueous systems. Here, above all disuccinimidyl carbonate, disuccinimidyl oxalate, glutaraldehyde, dimethylsuberimidate dihydrochloride or phenylene diisothiocyanate have to be mentioned as the reagents to be chosen. The reaction is carried out preferably under basic conditions, i.e. with the addition of diisopropylethylamine or NaCO<sub>3</sub>.

Principle:



The invention is further described by means of the figures which show:

Figure 1: generation of dendrimer structures after activation with acryloylchloride

Figure 2: generation of dendrimer structures after activation with 4-nitrophenylchloroformate

The invention is described in more detail by means of the examples.

**Example 1: Derivatization of a glass support**

It is started from aminated glass supports (40 x 48 mm) which is either commercially available or can be prepared according to the common methods of silanization. For activating the amino functions, the aminated glass supports are placed in 20 ml anhydrous dichloroethane with 200 mg 4-nitrophenyl chloroformate (1 mmol) and 171  $\mu$ l diisopropylethylamine (1 mmol) and swirled on a shaker for 2 hours. The BPB control test shows no blue staining. The glass supports are reacted overnight in 20 ml amine-free DMF with 223  $\mu$ l tetraethylene pentamine (1 mmol). The BPB control test shows a blue staining. For starting a new cycle, the glass supports are placed in 20 ml anhydrous dichloroethane with 200 mg 4-nitrophenyl chloroformate (1 mmol) and 171  $\mu$ l diisopropylethylamine (1 mmol) and swirled on a shaker for 2 hours. The BPB control test shows no blue staining. The glass supports are reacted overnight in 20 ml amine-free DMF with 140  $\mu$ l 1,4-bis(3-aminopropoxy)butane (1 mmol). The BPB test shows a blue staining.

The thus derivatized glass supports now have, on their surface, linkers to which the biopolymers (e.g. nucleic acids) can be linked.

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**Example 2: Derivatization of a glass support**

It is started from an aminated glass support (40 x 48 mm) which is either commercially available or can be prepared according to the common methods by means of silanization. For activating the amino functions, the aminated glass supports are placed in 20 ml anhydrous dichloroethane with 81  $\mu$ l acryloylchloride (1 mmol) and 171  $\mu$ l diisopropylethylamine (1 mmol) and swirled on a shaker for 2 hours. The BPB control test shows no blue staining. The glass supports are reacted for 2 nights in 20 ml amine-free DMF with 223  $\mu$ l tetraethylene pentamine (1 mmol). The BPB control test shows a blue staining. For starting a new cycle, the glass supports are placed in 20 ml anhydrous dichloroethane with 81  $\mu$ l acryloylchloride (1mmol) and 171  $\mu$ l diisopropylethylamine (1 mmol) and swirled for 2 nights on a shaker. The BPB control test shows no blue staining. The glass supports are reacted for 2 nights in 20 ml amine-free DMF with 140  $\mu$ l 1,4-bis(3-aminopropoxy)butane (1mmol). The BPB test shows a blue staining.

The thus derivatized glass supports then have, on their surface, linkers to which the biopolymers (e.g. nucleic acids) can be linked.

**Example 3: Derivatization of polypropylene membranes**

It is started from a purchasable hydrophilized polypropylene membrane (8 x 12 cm). For the purpose of activation, the membrane is reacted with 324  $\mu$ l acryloylchloride (4 mmol) and 684  $\mu$ l diisopropylethylamine (4 mmol) in 30 ml anhydrous dichloroethane on a shaker. After 2 hours, the reaction solution is decanted and washed several times carefully with dichloroethane and dried. The bromophenol blue test (BPB

test) shows no blue staining. The activated membrane is subsequently swirled for 2 days with 892 µl tetraethylene pentamine (4 mmol) in 30 ml amine-free DMF on a shaker. Then, the reaction solution is decanted and the membrane is washed carefully several times with DMF and then with dichloroethane and dried. The bromophenol blue test shows blue staining. For further activation the then aminated membrane is again reacted with 324 µl acryloylchloride (4 mmol) and 684 µl diisopropylethylamine (4 mmol) in 30 ml anhydrous dichloroethane on a shaker. After 2 hours, the reaction solution is decanted and washed several times carefully with dichloroethane and dried. The bromophenol blue test (BPB test) shows no blue staining. Thereafter, the activated membrane is swirled for 2 days with 560 µl 1,6-bis(3-aminopropoxy)butane (4 mmol) in 30 mol amine-free DMF on a shaker. Then, the reaction solution is decanted and the membrane is washed carefully several times with DMF and then with dichloroethane and dried. The bromophenol blue test (BPB test) shows blue staining.

**Example 4: Binding of nucleic acids to polypropylene membranes**

a) Binding by means of an activating agent

A polypropylene membrane derivatized according to Example 3 (8 x 12 cm) is incubated on a Whatman filter paper for 5 min. with a solution of 54 mg N-(3-dimethylaminopropyl)-N'-ethyl-carbodiimide hydrochloride and 24 ml N-methylimidazole in 10 ml water. The reaction solution is decanted. The thus activated still moist polypropylene membrane can then be used for covalent application of biopolymers (nucleic acids). After applying the biopolymers (e.g. by spotting robots

or nano plotter), the samples are fixed on the membrane by 2 h of incubation at 65°C; thereafter the membrane is washed carefully with water.

- b) Binding by means of electrostatic interaction under non-denaturing conditions

A polypropylene membrane derivatized in analogy to Example 3 (8 x 12 cm) is moistened with water on a filter paper. The moist membrane can then be used directly for the application of biopolymers. Having applied the biopolymers (e.g. by means of a spotting robot) the samples are fixed on the membrane by 2 hours of incubation at 65°C. Thereafter, the membrane is washed thoroughly with water.

As an alternative, the biopolymers can also be applied directly onto a dry membrane.

- c) Binding by means of electrostatic interaction under denaturing conditions

A polypropylene membrane derivatized in analogy to Example 3 (8 x 12 cm) is soaked on a filter paper with 10 ml denaturing buffer (0.5 M NaOH, 1.5 M NaCl). After 10 minutes of incubation, the supernatant of denaturing buffer is decanted. The still moist membrane can then be used for immobilizing biopolymers. Having applied the biopolymers (e.g. by means of a spotting robot), the membranes are dried overnight at room temperature. Thereafter, the membrane is washed thoroughly with water.

**Example 5:      Synthesis of terminal hydroxyl groups**

## a) Activation with chloroformic acid-4-nitrophenyl ester:

An amino-functionalized polypropylene membrane (omnitray size) according to Example 3 is swirled in 30 ml anhydrous dichloroethane with 400 mg chloroformic acid-4-nitrophenylester (2 mmol) and 342  $\mu$ l diisopropylethylamine (2 mmol) for 2 hours on a shaker. Washing is carried out 2 x for 5 minutes each with 50 ml dichloromethane and drying takes place in a nitrogen current. The thus activated PP membrane is then swirled with 200 ml 2-(2-aminoethoxy)ethanol (2 mmol) in 30 ml amine-free DMF for 30 min. on a shaker. The reaction solution is then decanted. After short washing with DMF, the PP membrane is again admixed with 200  $\mu$ l 2-(2-aminoethoxy)ethanol (2 mmol) in 30 ml amine-free DMF and swirled overnight on a shaker. The reaction solution is decanted and the PP membrane is washed 2 times for 5 minutes each with 50 ml DMF and then 2 times with 50 ml acetone and is dried.

## b) Activation with acryloylchloride:

An amino-functionalized polypropylene membrane (omnitray size) according to Example 3 is swirled in 30 ml anhydrous dichloroethane with 162  $\mu$ l acryloylchloride (2 mmol) and 342  $\mu$ l diisopropylethylamine (2 mmol) for 2 hours on a shaker. Washing is carried out 2 times for 5 minutes each with 50 ml dichloromethane and drying takes place in a nitrogen current. The thus activated PP membrane is then swirled with 200  $\mu$ l 2-(2-aminoethoxy)ethanol (2 mmol) in 30 ml amine-free DMF for 30 minutes on a shaker, whereupon the reaction solution is decanted. After short washing with DMF, the PP membrane is again admixed with 200  $\mu$ l 2-(2-aminoethoxy)ethanol (2 mmol) in 30 ml amine-free DMF and swirled overnight on a shaker. The reaction solution is

decanted and the PP membrane is washed 2 times for 5 minutes each with 50 ml DMF and then 2 times with 50 ml acetone and is dried.

**Example 6:      Synthesis of terminal carboxyl groups**

An amino-functionalized polypropylene membrane (omnitray size) according to Example 3 is reacted in 30 ml dry dichloroethane with 400 mg succinic anhydride (4 mmol), 1 ml pyridine (24 mmol) and 488 mg DMAP (4 mmol) overnight on a shaker. Subsequently, the reaction solution is decanted and the PP membrane is washed in succession with 30 ml dichloromethane, 30 ml acetone, 30 ml 1 % HCl, 2 x 30 ml water and 2 x with 30 ml acetone for 5 minutes each and is dried.

**Example 7:      Synthesis of terminal phosphate groups**

While cooling in an ice bath, 0.42 ml POCl<sub>3</sub> (4 mmol) is dissolved in 40 ml dry pyridine. Thereafter, 828 mg 1,2,4-triazole (12 mmol) are added. After 10 minutes, the hydroxyl-functionalized polypropylene membrane according to Example 4 is added and swirled for another 10 minutes with ice bath cooling on a shaker. Thereafter, the ice bath is removed and swirling is continued for 45 minutes. The reaction solution is decanted and carefully hydrolyzed with 50 ml of a mixture of triethylamine / dioxane / water. Washing is carried out carefully two times with 50 ml water, shortly with 50 ml 1 % HCl, 2 times with 50 ml water and 2 times for 5 minutes with 50 ml acetone and drying takes place in a nitrogen current.

**Example 8: Surface activation with disuccinimidyl carbonate**

7 amino-functionalized glass slides are swirled in 30 ml anhydrous acetonitrile with 256 mg disuccinimidyl carbonate (2 mmol) and 513 µl diisopropylethylamine (3 mmol) for 4 hours on a shaker. Washing is carried out with 50 ml acetonitrile, then with 50 ml dichloroethane and drying takes place in a nitrogen current.

**Example 9: Methylation of a glass support for improving the properties for the immobilization of biomolecules by electrostatic interaction**

7 items of derivatized glass supports (derivatized in analogy to Example 2) are incubated for 1.5 hours with a solution of 0.125 ml trifluoromethanesulfonic acid methyl ester (1.1 mmol) in 40 ml dichloroethane. Thereafter, thorough washing is carried out with dichloroethane and acetone and the supports are dried.

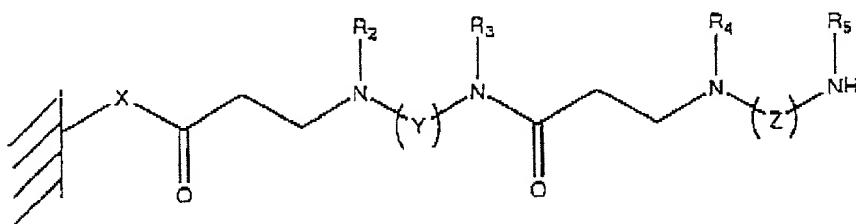
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**Claims**

- 1) A method of derivatizing carriers or supports, wherein a functional group is activated on a carrier or support surface by reaction with an activating reagent and subsequently reacted with an amine component.
- 2) The method according to claim 1, wherein the supports are selected from the group consisting of glass, sheets or films or membranes made from polypropylene, nylon, cellulose, cellulose derivatives (e.g. cellulose acetate, cellulose-mixed ester), polyether sulfones, polyamides, polyvinyl chloride, polyvinylidene fluoride, polyester, polyethylene or Teflon.
- 3) The method according to claim 1 or 2, wherein the functional group is an amine, hydroxyl, phosphate, carboxyl, carbonyl, thiol or amide group.
- 4) The method according to any one of claims 1 to 3, wherein the activating reagent is acryloylchloride, 4-nitrophenylchloroformate, carbonyl diimidazole, phenyl chloroformate, phosgene, disphosgene, triphosgene, EDC-(N-(3-dimethylaminopropyl)-N'-ethyl-carbodiimide hydrochloride), N,N'-diisopropyl carbodiimide, dicyclohexyl carbodiimide, disuccinimidyl carbonate, disuccinimidyl oxalate, dimethylsuberimidate dihydrochloride or phenylene diisothiocyanate.
- 5) The method according to any one of claims 1 to 4, wherein the amine component is selected from the group consisting of monoamines, bis-amines or polyamines.

- 6) The method according to claim 5, wherein the monoamine is 2-aminoethanol, 6-amino-1-hexanol, 2-(4-aminophenyl)ethanol, 5-amino-n-valeric acid, 2-(2-aminoethoxy)ethanol or 3-amino-1,2-propanediol.
- 7) The method according to claim 5, wherein the bis-amine is 1,4-bis(3-aminopropoxy)butane, O,O'-bis(2-aminopropyl)polyethylene glycol 500 (= Jeffamine 500), O,O'-bis(2-aminopropyl)polyethylene glycol 130 (= Jeffamine 130), 4,7,10-trioxa-1,13-tridecaneamine or ethylene diamine.
- 8) The method according to claim 5, wherein the polyamine is tetraethylene pentamine, spermine, spermidine, 4,7,10-trioxa-1,13-tridecanediamine or 4-aminomethyl-1,8-octanediamine.
- 9) The method according to any one of claims 1 to 8, wherein the steps of the reaction with an activating reagent and an amine component are carried out several times.
- 10) The method according to claim 9, wherein dendrimer structures are built up on the support surface.
- 11) The method according to any one of the preceding claims, wherein positive charge is built up in controlled fashion on the support surface.
- 12) The method according to any one of claims 1 to 11, wherein the support surface derivatized according to any one of claims 1 to 11 is additionally activated prior to the attachment of biopolymers.

- 13) The method according to claim 12, wherein disuccinimidyl carbonate, disuccinimidyl oxalate, glutaraldehyde, dimethylsuberimidate dihydrochloride or phenylene diisothiocyanate are used as activation agent.
- 14) A support suitable for the attachment of biopolymers, wherein the surface of the support includes linkers having the following structure:



X = O, NHR<sub>1</sub>

Y, Z = may be equal or different and be selected from

- (CH<sub>2</sub>)<sub>n</sub>-

- (CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>-CH<sub>2</sub>CH<sub>2</sub>

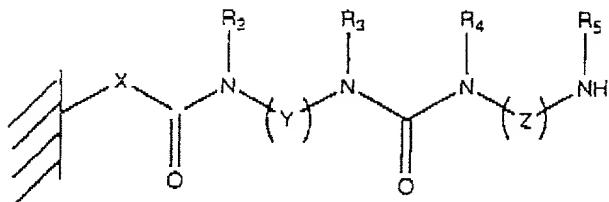
- (CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>-CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>-, etc.

R<sub>1</sub>-R<sub>5</sub> = may be equal or different and be selected from a straight-chain or branched alkyl residue having 1 to 30 carbon atoms, straight-chain or branched alkenyl residue having 2 to 30 carbon atoms, monocyclic or polycyclic alkyl residue having 3 to 30 carbon atoms or monocyclic or polycyclic alkenyl residue having 4 to 30 carbon atoms or monocyclic or polycyclic residue having 6 to 30 carbon atoms, the residues being optionally substituted by one or several

substituents (e.g. OH, carboxyl, carbonyl, phosphate)

n = 1 to 50

or



X = O, NHR<sub>1</sub>

Y, Z = may be equal or different and be selected from

- (CH<sub>2</sub>)<sub>n</sub>-

- (CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>-CH<sub>2</sub>CH<sub>2</sub>

- (CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>O)<sub>n</sub>-CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>-, etc.

R<sub>1</sub>-R<sub>5</sub> = may be equal or different and be selected from a straight-chain or branched alkyl residue having 1 to 30 carbon atoms, straight-chain or branched alkenyl residue having 2 to 30 carbon atoms, monocyclic or polycyclic alkyl residue having 3 to 30 carbon atoms or monocyclic or polycyclic alkenyl residue having 4 to 30 carbon atoms or monocyclic or polycyclic residue having 6 to 30 carbon atoms, the residues being optionally substituted by one or several substituents (e.g. OH, carboxyl, carbonyl, phosphate)

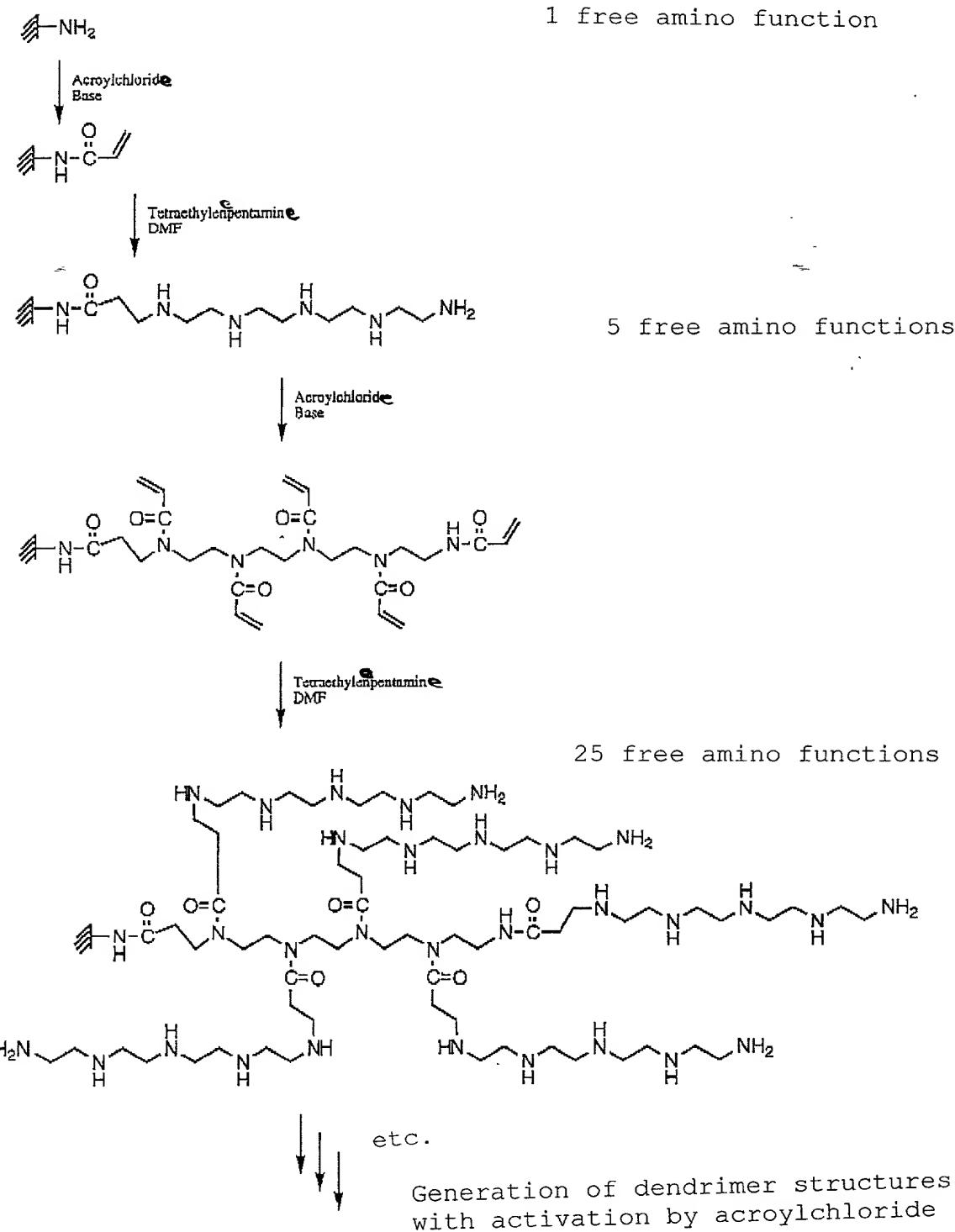
n = 1 to 50

- 15) The support suitable for the attachment of biopolymers, which includes linkers in the form of dendrimer structures on its surface.
- 16) Use of a support produced according to any one of claims 1 to 11 or a support according to claim 14 or 15 for the attachment of biopolymers.
- 17) Use according to claim 16, wherein the biopolymers are selected from the group consisting of DNA, RNA, nucleotide analogs, peptides, proteins or antibodies.

**Abstract of the Disclosure**

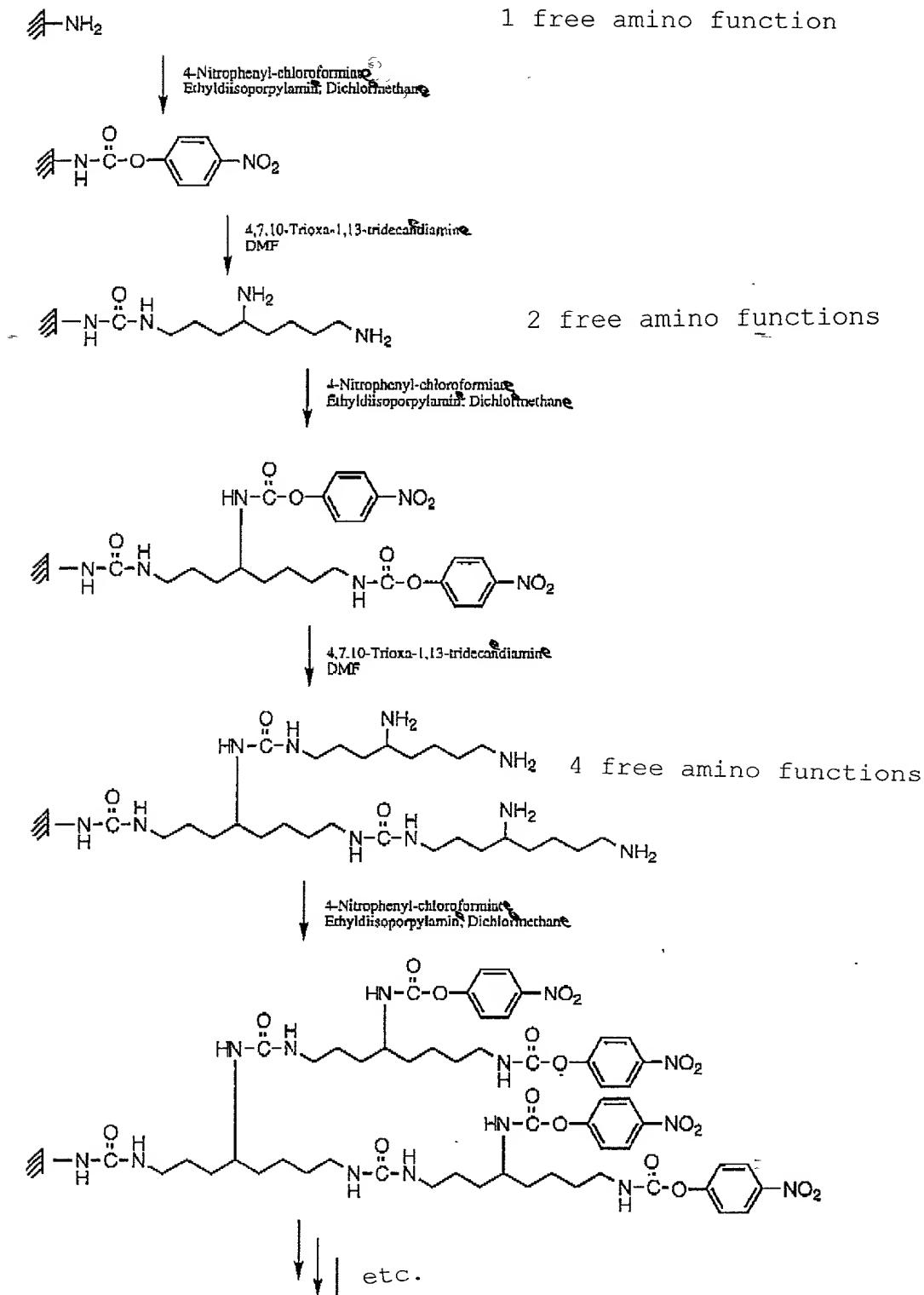
The present invention relates to a process for derivatizing supports or carriers, wherein a functional group is activated on a support surface by reaction with an activating reagent and then reacted with an amine component. The invention also relates to a support with a dendrimer structure on its surface and to the use of a support that has been produced according to the invention for binding biopolymers.

1/2



[ Fig.1 ]

2/2



[ Fig.2 ]

Generation of dendrimer structures  
with activation by 4-nitrophenyl chloroformate

**Combined Declaration and Power of Attorney for Patent Application**Docket Number: 03528.0131.PCUS00

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter that is claimed and for which a patent is sought on the invention entitled DERIVATIZATION OF SUPPORT SURFACES, the specification of which is attached hereto unless the following box is checked:

was filed on May 18, 2001;  
as United States Application Number or PCT International Application Number 09/856,341; and  
was amended on May 18, 2001 (if applicable).

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information that is material to patentability as defined in 37 C.F.R. § 1.56.

I hereby claim foreign priority benefits under 35 U.S.C. § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT international application, which designated at least one country other than the United States listed below, and have also identified below any foreign application for patent or inventor's certificate, or PCT international application having a filing date before that of the application on which priority is claimed.

Prior Foreign Application(s)	Priority Claimed
<u>PCT/DE99/03692</u> (Application No.)	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No
<u>PCT</u> (Country)	<u>November 17, 1999</u> (Day/Month/Year Filed)
<u>DE198 53 242.3</u> (Application No.)	<input checked="" type="checkbox"/> Yes <input type="checkbox"/> No
<u>DE</u> (Country)	<u>November 18, 1998</u> (Day/Month/Year Filed)

I hereby claim the benefit under 35 U.S.C. § 119(e) of any United States provisional application(s) listed below.

(Application No.)                          (Filing Date)

(Application No.)                          (Filing Date)

I hereby claim the benefit under 35 U.S.C. § 120 of any United States application(s), or under § 365(c) of any PCT international application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States or PCT international application in the manner provided by the first paragraph of 35 U.S.C. § 112, I acknowledge the duty to disclose information that is material to patentability as defined in 37 C.F.R. § 1.56 that became available between the filing date of the prior application and the national or PCT international filing date of this application.

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(Application No.)	(Filing Date)	(Status - patented, pending, abandoned)
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(Application No.)	(Filing Date)	(Status - patented, pending, abandoned)
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I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith:

Jason C. Abair, Reg. No. 44,007  
Ethan B. Andelman, Reg. No. P-48,997  
William D. Belanger, Reg. No. 40,509  
Michael J. Bell, Reg. No. 39,604  
Luisa Bigornia, Reg. No. 45,974  
Andrew S. Brenc, Reg. No. 45,534  
Celine T. Callahan, Reg. No. 34,301  
Jenny W. Chen, Reg. No. 44,604  
Robin C. Chiang, Reg. No. 46,619  
June E. Cohan, Reg. No. 43,741  
Chris Comuntzis, Reg. No. 31,097  
Thomas E. Coverstone, Reg. No. 36,492  
Ross E. Davidson, Reg. No. 41,698  
James F. Davis, Reg. No. 21,072  
Thomas M. Dunham, Reg. No. 39,965  
Joel Mark Freed, Reg. No. 25,101  
Darren J. Gold, Reg. No. 47,599  
Alan M. Grimaldi, Reg. No. 26,599  
J. Jay Giuliano, Reg. No. 41,810  
Albert P. Halluin, Reg. No. 25,227  
Leslie L. Jacobs, Jr., Reg. No. 40,659  
Derek J. Jardieu, Reg. No. 44,483

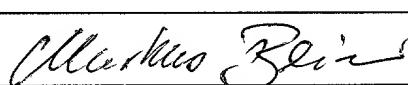
Christopher L. Kelley, Reg. No. 42,714  
Brian S.Y. Kim, Reg. No. 41,114  
Viola T. Kung, Reg. No. 41,131  
Robert C. Laurenson, Reg. No. 34,206  
Joseph P. Lavelle, Reg. No. 31,036  
Don F. Livornese, Reg. No. 32,040  
David R. Marsh, Reg. No. 41,408  
Christopher A. Mathews, Reg. No. 35,944  
Matthew J. Moore, Reg. No. 42,012  
Andrew Y. Piatnicia, Reg. No. 40,772  
M. Todd Rands, Reg. No. 46,249  
William C. Rookridge, Reg. No. 31,791  
Michael J. Stimson, Reg. No. 45,429  
Jennifer A. Tipsord, Reg. No. 40,205  
William K. West, Reg. No. 22,057  
Justin A. White, Reg. No. P-48,883  
Adam K. Whiting, Reg. No. 44,400  
Jayna R. Whitt, Reg. No. 47,175  
Karen K. Wong, Reg. No. 44,409  
Robert J. Worrall, Reg. No. 37,969  
Wallace Wu, Reg. No. 45,380  
Matthew S. Zises, Reg. No. 47,246

Send Correspondence to:

**Albert P. Halluin**  
**HOWREY SIMON ARNOLD & WHITE, LLP**  
301 Ravenswood Avenue  
Box No. 34  
Menlo Park, CA 94025  
Facsimile: (650) 463-8400

Direct Telephone Calls to: (650) 463-8109

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full name of sole or first inventor <u>Markus BEIER</u>	
Inventor's signature 	Date <u>10 Aug 01</u>
Residence Werderstrasse 42A, D-69120 Heidelberg	<u>DEX</u>
Citizenship DE	
Post Office Address Werderstrasse 42A, D-69120 Heidelberg	

(Supply similar information and signature for subsequent joint inventors, if any)